

Didymosphenia geminata [简体中文](#) [正體中文](#)

System: Freshwater

Kingdom	Phylum	Class	Order	Family
Plantae	Bacillariophyta	Bacillariophyceae	Cymbellales	Gomphonemataceae

Common name didymo (English, New Zealand), rock snot (English, New Zealand)

Synonym *Echinella geminata* , (Lyngbye, 1819)
Gomphonema geminatum , (Lyngbye) Agardh 1824

Similar species *Cymbella* spp., *Gomphoneis* spp., *Gomphonema* spp.

Summary *Didymosphenia geminata* is a freshwater diatom which has historically been found in cool, oligotrophic waters of northern Europe and northern North America. Since the mid-1980s, it has begun to take on the characteristics of an invasive species in both its native range and introduced regions (New Zealand). It can form massive blooms which have a range of adverse effects on freshwater ecosystems, as well as human and economic effects.



[view this species on IUCN Red List](#)

Species Description

Didymosphenia geminata is a larger freshwater diatom (Kilroy, 2004). It can form massive blooms on the bottom of streams, rivers, and rarely in lakes. It is both epilithic (attaching to stones) and epiphytic (attaching to plants) (Round *et al.* 1990; in Kilroy, 2004), attaching itself by stalks, and it can form a thick brown layer that smothers rocks, submerged plants and other materials. It forms flowing 'rats tails' that can turn white at their ends and look similar to tissue paper. Although the algae appear slimy, it feels like wet cotton wool. (Biosecurity NZ, 2005). The mats consist mainly of the stalks, which contain no chlorophyll, and give the mat a pale yellow-brown to white colour (Ministry of Water, Land and Air Protection, undated).

D. geminata is made up of cells that cannot be seen with the naked eye until large colonies form. It only needs a single cell to be transported for the algae to spread (Biosecurity NZ, 2005). These cells are distinguished by their large, triundulate frustule, shaped like a curved bottle, and prominent striae (regular lines of holes starting at the centre line of the valve faces) which are radially arranged and variable in length at the centre (Kilroy, 2004).

D. geminata can be distinguished from other species of algae on the basis of: **Colour** *D. geminata* is beige/brown/white but not green. **Touch** Although it looks slimy it doesn't feel slimy, but rather spongy and scratchy like cotton wool. **Odour** Live *D. geminata* has no distinctive odour. **Strength** *D. geminata* is strongly attached to river stones and does not fall apart when rubbed between your fingers (Biosecurity NZ, 2005).

Notes

It is unknown what has changed to allow *Didymosphenia geminata* to take on the characteristics of an invasive species, but connections between the features of the diatom (cell size, stalk composition), other organisms (mayflies, stoneflies, midge larvae) and the physical environment (stream flow, ultraviolet light, temperature, sediment) may play a role in explaining the dense growths of this diatom (Spaulding, 2005).

One current theory suggests that the increasing occurrence of *Didymosphenia geminata* may be attributed to a genetic variant which has broader tolerances than the original species (Kilroy, 2004). It has also been suggested that *Didymosphenia* may proliferate because of increased exposure to ultraviolet (UV) radiation. This could occur because the UV reduces the grazer populations that normally limit accumulation of *Didymosphenia*, or because *Didymosphenia* outcompetes other algal species under increased UV conditions. Under this theory, warmer winters and reduced flows may favour growth of *Didymosphenia* - in other words, climate change may be linked to the current range expansion being observed for *Didymosphenia* (Kilroy, 2004).

There appears to be some confusion as to the number of species contained within the genus *Didymosphenia*, with various species other than *Didymosphenia geminata* being identified in the literature.

D. dentata was recorded from Lake Baikal and is described as having a valve exterior with occluded areolae, prominent marginal spines, dilated proximal raphe ends and distal ends that are deflected at 90 degree angles in the same direction. Ridges or flanges spiral around each spine. Stigmata are lacking. Internally the central nodule, raphe sternum and helictoglossae are prominent. *D. dentata* is said to differ from *D. geminata* in valve shape, presence of marginal spines and lack of stigmata. Recent suggestions that the two species are conspecific are not supported by observations presented in Kociolek *et al* (2000).

Metzeltin and Lange-Bertalot (1995), in their taxonomic evaluation of *Didymosphenia* identified three morphotypes of *D. geminata*: *geminata sensu stricto*, *capitata* and *subcapitata*. They also recognised another four species: *D. siberica* (Grun.) M. Schmidt, *D. curvata* (Skv. and Meyer), *D. clavaherculis* (Ehr.) and *D. pumila* nov. spec. These can be differentiated by the structure of the areolae, which is profoundly different in *D. geminata* and *D. clavaherculis* compared to *D. siberica*, *D. curvata* and *D. pumila*.

Uses

Didymosphenia geminata has been used as an indicator of oligotrophic conditions (Sahin, 2000).

Habitat Description

Didymosphenia geminata is found in freshwater rivers, streams and lakes. It occurs particularly in oligotrophic, clear-water montane or northern boreal streams (Kilroy, 2004). *D. geminata* thrives in clear, shallow, warm and nutrient-poor water, and is influenced annually by weather and rainfall patterns. High light and stable flow conditions are also favourable for *D. geminata* (Kilroy, 2004). It is normally found in water with pH=7, but appears to do well where there is some calcium (E.Y. Haworth, pers. comm., December 2004; in Kilroy, 2004). Stable flow and a stable substrate are probably required for the initial attachment to the substrate. Most *Didymo* blooms reported occur in lake-fed rivers or in regulated rivers (below dams), ie., in freshwater systems characterized by relatively stable flows. Once a colony is established, fast currents are likely to enhance growth by promoting transfer of nutrients to the cells at the mat surface (Kilroy, 2004). An important habitat requirement for *D. geminata* is high light levels (Kawecka and Sanecki, 2003; in Kilroy, 2004). In British Columbia, *D. geminata* was found in depths from 10cm to 2m, with the heaviest biomass occurring in high-light areas (Kilroy, 2004). Detailed analysis of the blooms suggested that the distribution of *Didymosphenia* blooms may also be related to geological factors and temperature (Sherbot and Bothwell, 1993; in Kilroy, 2004).

Reproduction

Didymosphenia geminata populations, like those of other diatoms, grow by vegetative cell division. The two valves of the cell each form a new valve which fits inside the original one, thus causing a gradual average reduction in cell size in a population (Round *et al.*, 1990; in Kilroy, 2004). Each branch point in the stalk represents a vegetative cell division. Most diatoms also undergo sexual reproduction at some stage, which restores the size of the cells to their maximum (Kilroy, 2004).

Nutrition

Didymosphenia geminata is a primary producer that contains chlorophyll.

General Impacts

Since the mid-1980s, *D. geminata* appears to have been gradually expanding its geographical range in North America and Europe. There have also been reports of it occurring in high abundances in areas where it has previously only been known in low levels (Whitton and Crisp, 1984; Heuff and Horkan, 1984; Skulberg and Lillehammer, 1984; in Kilroy, 2004). *D. geminata* can form massive blooms which can adversely affect freshwater fish, plant and invertebrate species, e.g. by reducing the number of suitable habitats and excluding the growth of other diatoms (Biosecurity New Zealand, 2005).

Kilroy (2004) assesses the impacts of the spread of *D. geminata* in New Zealand. Some of the impacts include: an effect on the tourism industry, significant aesthetic effects, alteration in invertebrate communities, minor health effects and the possibility of spread to other rivers.

For more details follow this link [Kilroy, 2004](#)

Concerns raised by the Environment Protection Division of the Ministry of Environment, Government of British Columbia of possible impacts of *D. geminata* blooms include reduction in the rearing habitat for salmonid species, alterations in the species composition and populations of invertebrates, restriction of water flow and depletion of dissolved oxygen in the water due to decomposition of algal mats (EPD, undated).

Potential impacts of the spread of *D. geminata* discussed by the MDDEP-MRNF report include a possible alteration of the species composition of benthic invertebrate communities which could affect fish populations; alteration in streamflow and an effect on recreational activities (MDDEP-MRNF, 2007).

An electric fishing survey was carried out by the Ministère des Ressources Naturelles et de la Faune (MRNF), in the Matapedia river in Quebec, in early September 2006. Large *D. geminata* blooms had been observed in the river. The MRNF was not able to measure impacts on the abundance of juvenile salmon. The MDDEP-MRNF (2007) states that a similar observation had been made by fisheries experts and managers from France, Iceland, Ireland, Scotland, Finland and Norway. No impacts had been recorded on either adults or juveniles of Atlantic salmon or any other salmonid species.

Management Info

Cultural: Following the discovery of *D. geminata* in Southland, Biosecurity New Zealand released the following advice for containment of the algae:

1. Before leaving the river, remove obvious clumps of algae, taking care to search for hidden clumps which may be obscured within compartments. Leave clumps at affected site. After leaving affected area, if you find clumps, do not wash clumps down drains. Treat them chemically or by drying as below.

2. Soak and scrub all items for at least one minute in hot (60°C) water, a 2 percent solution of household bleach or a 5 percent solution of salt, nappy cleaner, antiseptic hand cleaner or dishwashing detergent.

Note: For a 2 percent solution, add 200 mls to a ten litre bucket and fill with water. For a 5 percent solution, add 500 mls (two heavy cups) to a ten litre bucket and fill with water.

If cleaning using the approved chemical methods in step two is not feasible, alternatively the **infected equipment must be COMPLETELY dry for at least two days before it can be safely used in unaffected waters**. Equipment that remains damp or could have pockets of trapped moisture after use will require longer drying times because the 48 hour drying time begins after dampness and trapped moisture have evaporated. Some equipment may never reach complete dryness depending upon how and where it is stored (shoes, waders, life vests, wetsuits, spray skirts, jet boat intakes, tyres, etc.) and therefore should always be chemically treated.

Under no circumstances should fish, plants or other items be moved from an affected waterway to an unaffected waterway.

Pathway

The arrival of *D. geminata* in New Zealand in 2004 indicates that it most likely arrived via human-assisted means, for example on footwear, fishing equipment, boats, etc. (Kilroy, 2004).

Principal source: Kilroy, C. November 2004. [A new alien diatom, *Didymosphenia geminata* \(Lyngbye\) Schmidt: its biology, distribution, effects and potential risks for New Zealand fresh waters.](#)

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ALIEN RANGE

[2] CANADA	[1] CHILE
[5] CHINA	[1] HUNGARY
[1] ICELAND	[3] IRELAND
[1] KAZAKHSTAN	[1] KYRGYZSTAN
[1] MONGOLIA	[1] NEW ZEALAND
[1] NORWAY	[1] PAKISTAN
[14] POLAND	[2] ROMANIA
[7] RUSSIAN FEDERATION	[1] SVALBARD AND JAN MAYEN
[2] SWEDEN	[10] TURKEY
[3] UKRAINE	[5] UNITED KINGDOM
[8] UNITED STATES	

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